

## Role of tyrosine phosphatase in the modulation of pulmonary vascular tone

X.L. Huang\*, D. El Kebir\*, A.S. De Buys Roessingh\*, J.C. Schneider\*, L. Jacob#, J.C. Mercier<sup>†</sup>, J. Dall'Ava-Santucci\*, A.T. Dinh-Xuan\*

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**ABSTRACT:** In the vascular system, synthesis of the potent vasodilator nitric oxide (NO) is tightly regulated by the constitutively expressed endothelial NO synthase (eNOS). Activity of eNOS is controlled by  $Ca^{2+}$ /calmodulin and various seryl/threonyl protein kinases. Less is known about the importance of phosphorylation and dephosphorylation of tyrosyl residues. Therefore the role of tyrosine phosphatase on the modulation of isolated rat pulmonary artery tone has been assessed. Inhibition of tyrosine phosphatase by sodium orthovanadate (SOV,  $1 \times 10^{-6}$  M) significantly: 1) increased phenylephrine-induced vasoconstriction and 2) decreased endothelium-dependent relaxation to acetylcholine, but had no effect on endothelium-independent relaxation to the NO donor, sodium nitroprusside. In phenylephrine-precontracted pulmonary arterial rings, SOV ( $1 \times 10^{-7}$ – $1 \times 10^{-5}$  M) had no effect on vascular tone but significantly relaxed rings which were pretreated with the NO-synthase inhibitor, *N*<sup>ω</sup>-nitro-L-arginine-methyl ester (L-NAME). SOV-induced relaxation in the presence of L-NAME was, however, abolished by glibenclamide.

In conclusion, inhibition of tyrosine phosphatase altered pulmonary vascular tone by increasing vasoconstrictor response to phenylephrine and decreasing endothelium-dependent relaxation to acetylcholine. Furthermore, the tyrosine phosphatase inhibitor, sodium orthovanadate, exhibited original vasodilator properties which were only observed when nitric oxide synthesis was inhibited. Thus a new pathway involving the inhibitory effect of nitric oxide on a glibenclamide-sensitive diffusible relaxing factor, that might play an important role in the control of pulmonary vascular tone is described.

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\*Service de Physiologie-Explorations Fonctionnelles and #Laboratoire de Biologie Cellulaire, Centre Hospitalier Universitaire, Cochin Assistance Publique-Hôpitaux de Paris, Université Paris V, 75679 Paris cedex 14. <sup>†</sup>Service de Réanimation Pédiatrique Hôpital Robert Debré, Assistance Publique-Hôpitaux de Paris, Université Paris VII, France.

Correspondence: A.T. Dinh-Xuan, Service de Physiologie-Explorations Fonctionnelles, Centre Hospitalier Universitaire Cochin, Assistance Publique-Hôpitaux de Paris, Université Paris V, 27 rue du faubourg Saint-Jacques, 75679 Paris cedex 14, France.  
Fax: 33 158412345  
E-mail: anh-tuan.dinh-xuan@cch.ap-hop-paris.fr

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Pulmonary vascular tone is modulated by a variety of endothelium-derived relaxing factors including prostacyclin, nitric oxide (NO) [1, 2], and the endothelium-derived hyperpolarizing factors (EDHF) [2, 3]. NO is one of the most potent vasodilators known to date. Its synthesis is tightly regulated by intracellular free  $Ca^{2+}$  and the  $Ca^{2+}$ /calmodulin complex that, in turn, stimulate the constitutive isoform of endothelial NO synthase (eNOS). Alternatively, phosphorylation on seryl and threonyl residues by various seryl/threonyl kinases is also thought to modulate eNOS activity [4, 5].

Shear stress is the major physiological factor modulating pulmonary vascular tone in health and disease. The underlying mechanism most likely involves synthesis and release of NO from pulmonary endothelial cells. Recent evidence suggests that endothelial production of NO is increased in response to shear stress through tyrosine phosphorylation *via* a calcium-independent pathway in the systemic circulation [4, 6]. Contradictory studies alternatively

suggest that tyrosine phosphorylation of eNOS might also reduce its activity [7]. Whether or not alteration of phosphorylation and/or dephosphorylation of tyrosyl residues of eNOS affects its activity in the pulmonary artery has yet to be investigated.

In various vascular beds, EDHF causes relaxation through vascular smooth muscle hyperpolarization [3]. The identity of EDHF is still under debate. Early evidence has suggested that adenosine triphosphate (ATP)-dependent potassium ( $K_{ATP}$ ) channels are activated by EDHF [8–10], whereas recent evidence further suggests that EDHF is a product of cytochrome P-450 enzymes [11, 12] which cause relaxation by activating large conductance calcium-activated potassium ( $K_{Ca}$ ) channels [11–13]. The physiological role of EDHF in pulmonary vessels is still uncertain, although there is circumstantial evidence to suggest that EDHF might play an important role in transitional pulmonary circulation [14].

The aim of this study was to further elucidate the role of tyrosine phosphorylation in NO and EDHF

signalling pathways in the pulmonary circulation. The authors have more specifically studied the effect of sodium orthovanadate (SOV), a tyrosine phosphatase inhibitor, which has been demonstrated to activate NOS through tyrosine phosphorylation-dependent mechanisms in the systemic circulation [4, 15]. It was found that SOV significantly altered pulmonary vascular tone by reducing NO-mediated pulmonary vasodilatation and increasing the vasodilator effects of EDHF.

### Materials and methods

Male Sprague-Dawley rats (250–275 g) were anaesthetized intraperitoneally with thiopental sodium (80 mg·kg<sup>-1</sup>). The pulmonary arteries (PA) were immediately dissected and placed in cold Krebs solution: NaCl 118, KCl 5.9, MgSO<sub>4</sub> 1.2, CaCl<sub>2</sub> 2.5, NaH<sub>2</sub>PCO<sub>4</sub> 1.2, glucose 5.6, NaHCO<sub>3</sub> 25.5 (mM). The arteries were cleaned of perivascular tissue and cut into rings 2.5 mm in length. The rings were suspended between two wire hooks. One of the hooks was fixed to a support in an isolated organ bath containing 20 mL of Krebs solution maintained at 37°C and gassed with a mixture of 95% oxygen (O<sub>2</sub>) and 5% carbon dioxide (CO<sub>2</sub>). Each vascular segment was set at a resting tension for optimal length development ranging from 0.7–1.0 g. The rings were then allowed to equilibrate for 60 min. After equilibration, three studies were performed.

#### Experimental studies

After contraction with phenylephrine (1×10<sup>-7</sup>–1×10<sup>-6</sup> M) to obtain a stable plateau of tension, the rings were challenged with increasing concentrations of acetylcholine (1×10<sup>-7</sup>–1×10<sup>-5</sup> M). Sodium nitroprusside, a NO donor (1×10<sup>-4</sup> M), was added at the end of the experiments to maximally relax pulmonary vascular smooth muscle. PA rings were divided into two groups, one group were treated with SOV (1×10<sup>-6</sup> M) for 15 min, whereas the other group were treated with the solvent (distilled water) and served as controls. SOV was first added 15 min before contraction to phenylephrine to study the effects of SOV on phenylephrine-induced contraction.

In a separate set of experiments, SOV was added after the plateau of contraction to phenylephrine was reached, incubation with SOV then lasted for 15 min before endothelium-dependent relaxation to acetylcholine (1×10<sup>-7</sup>–1×10<sup>-5</sup> M) and endothelium-independent relaxation to the NO donor, sodium nitroprusside (1×10<sup>-4</sup> M), were assessed.

In a third set of experiments, the effects of increasing concentrations of SOV (1×10<sup>-7</sup>–1×10<sup>-5</sup> M) on phenylephrine-precontracted pulmonary vascular rings was assessed. The rings were divided into three groups: 1) untreated; 2) treated with N<sup>ω</sup>-nitro-L-arginine methyl ester (L-NAME) (1×10<sup>-3</sup> M); and 3) treated with L-NAME (1×10<sup>-3</sup> M) and glibenclamide (1×10<sup>-5</sup> M).

#### Drugs

All drugs were purchased from Sigma (Saint Quentin-Fallavier, France). SOV, phenylephrine, acetylcholine, sodium nitroprusside and L-NAME, were dissolved in distilled water, whereas glibenclamide was dissolved in dimethyl sulphoxide which, by itself had no effect on tone of isolated pulmonary vessels.

#### Data analysis

The magnitude of contraction by phenylephrine was expressed in mg (mean±SEM), n is the number of pulmonary vascular rings. Relaxation and contraction were expressed as the percentage decrease or increase in tone from the plateau of precontraction obtained with phenylephrine (1×10<sup>-6</sup> M). The mean±SEM of each set of experimental data were used to construct the concentration-response curves to various pharmacological substances. Statistical analysis was performed using the nonparametric Wilcoxon signed-rank test to compare results obtained on rings with and without endothelium that were challenged with various inhibitors or agonists and their controls. Values were considered statistically significant when p<0.05.

### Results

#### *Sodium orthovanadate increased phenylephrine-induced contraction*

SOV (1×10<sup>-6</sup> M), by itself, had no contractile effect on basal tone but significantly increased the magnitude of contraction in all PA rings (n=10) in response to phenylephrine (1×10<sup>-7</sup> M) (614±78 mg *versus* 745±90 mg; p<0.001) and 1×10<sup>-6</sup> M (767±102 mg *versus* 960±123 mg; p<0.0001).

In PA rings without endothelium, SOV (1×10<sup>-6</sup> M), also significantly affected the contractile responses to phenylephrine, increasing maximal tension from 851±168 mg to 1010±193 mg (in control and treated rings, respectively; p<0.01).

#### *Sodium orthovanadate reduced endothelium-dependent relaxation to acetylcholine*

In PA rings precontracted with phenylephrine (1×10<sup>-6</sup> M) pretreatment with SOV (1×10<sup>-6</sup> M) significantly decreased endothelium-dependent relaxation to acetylcholine (maximal relaxation 48±9% in treated PA rings *versus* 75±8% in controls, p<0.05) (fig. 1).

#### *Endothelium-dependent relaxation in response to sodium orthovanadate*

Increasing concentrations of SOV (1×10<sup>-7</sup>–1×10<sup>-5</sup> M) had no significant relaxing effect on PA

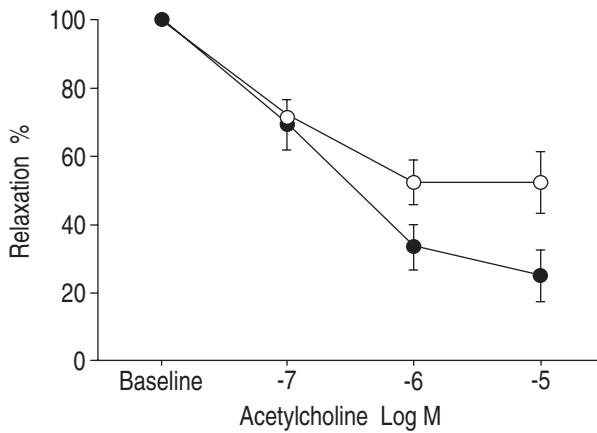


Fig. 1.—Effect of sodium orthovanadate (SOV) on acetylcholine-induced relaxation. Cumulative concentration-response curves for acetylcholine were made in the rings pretreated with SOV and in control rings. Relaxation was presented as percentage of precontraction with phenylephrine ( $1 \times 10^{-6}$  M). Data was expressed as mean  $\pm$  SEM. n=8 each. ●: control; ○: with SOV ( $1 \times 10^{-6}$  M).

rings precontracted with phenylephrine ( $1 \times 10^{-6}$  M) (fig. 2). However, pretreatment with L-NAME ( $1 \times 10^{-3}$  M) caused relaxation of PA rings challenged with increasing concentrations of SOV ( $21 \pm 5\%$  versus  $3 \pm 2\%$  for  $1 \times 10^{-7}$  M,  $28 \pm 7\%$  versus  $5 \pm 3\%$  for  $1 \times 10^{-6}$  M and  $30 \pm 8\%$  versus  $6 \pm 4\%$  for  $1 \times 10^{-5}$  M,  $p < 0.05$  respectively). L-NAME-induced endothelium-dependent relaxation to SOV was completely abolished by glibenclamide ( $1 \times 10^{-5}$  M) (fig. 2).

In PA rings without endothelium, SOV ( $1 \times 10^{-7}$ – $1 \times 10^{-5}$  M) similarly failed to induce any relaxing effect on PA rings precontracted with phenylephrine ( $1 \times 10^{-6}$  M). However, unlike PA rings with endothelium, neither L-NAME nor glibenclamide significantly

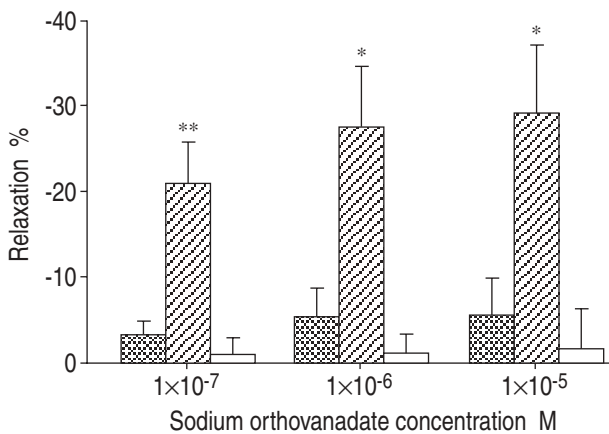


Fig. 2.—Effect of sodium orthovanadate (SOV) on pulmonary arterial rings with endothelium in the presence of: ▨: *N*<sup>ω</sup>-nitro-L-arginine-methyl ester (L-NAME) ( $1 \times 10^{-3}$  M, n=8); and □: a combination of L-NAME ( $1 \times 10^{-3}$  M) and glibenclamide ( $1 \times 10^{-5}$  M, n=7); ■: control, n=8. Relaxation in response to SOV ( $1 \times 10^{-7}$ ,  $1 \times 10^{-6}$  and  $1 \times 10^{-5}$  M) was expressed as percentage from precontraction to phenylephrine ( $1 \times 10^{-6}$  M). Data was expressed as mean  $\pm$  SEM of n experiments. \*:  $p < 0.05$ ; \*\*:  $p < 0.01$ .

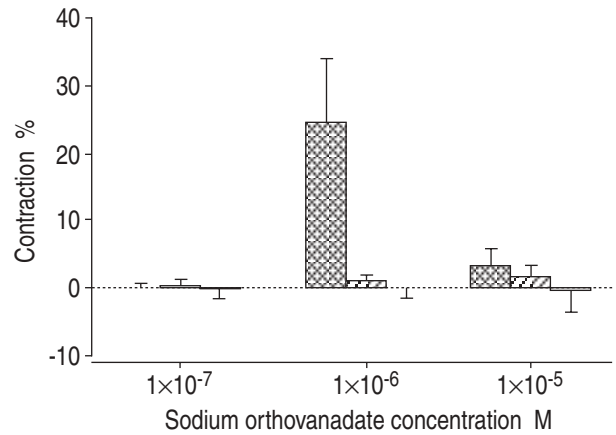


Fig. 3.—Effect of sodium orthovanadate (SOV) on pulmonary arterial rings without endothelium in the presence of: ▨: *N*<sup>ω</sup>-nitro-L-arginine-methyl ester (L-NAME) ( $1 \times 10^{-3}$  M); and □: a combination of L-NAME ( $1 \times 10^{-3}$  M) and glibenclamide ( $1 \times 10^{-5}$  M); ■: control. Response to SOV ( $1 \times 10^{-7}$ ,  $1 \times 10^{-6}$  and  $1 \times 10^{-5}$  M) was expressed as percentage from precontraction to phenylephrine ( $1 \times 10^{-6}$  M). Vasoconstriction was represented by positive values whereas vasorelaxation was represented by negative values. n=4. Data was expressed as mean  $\pm$  SEM of n experiments.

altered the lack of vasorelaxant effects of SOV in PA rings without endothelium (fig. 3).

*Effect of sodium orthovanadate on endothelium-independent relaxation to sodium nitroprusside*

In PA rings precontracted with phenylephrine ( $1 \times 10^{-6}$  M), SOV ( $1 \times 10^{-6}$  M) had no effect on endothelium-independent relaxation to the NO donor sodium nitroprusside.

**Discussion**

It has been demonstrated that inhibition of tyrosine phosphatase by SOV significantly altered pulmonary vascular tone by increasing vasoconstrictor response to phenylephrine and decreasing the endothelium-dependent vasodilator response to acetylcholine. As the increase in vasoconstrictor response to phenylephrine was also present in PA rings without endothelium, the procontractile effect of SOV would be best explained by a direct action on pulmonary vascular smooth muscle rather than putative release of endothelial-derived vasoconstrictors.

As reported previously in systemic circulation, inhibition of tyrosine phosphatase by SOV markedly increased vasoconstrictor responses to phenylephrine [16, 17]. This observation is consistent with the fact that phosphorylation of tyrosyl residues by tyrosine kinase is implicated in the mechanisms leading to constriction of vascular smooth muscle [18–22]. Vasoconstriction occurs as a result of activation of myosin light chain kinase (MLCK) that, in turn, phosphorylates myosin light chain, thereby enabling formation of actin-myosin cross bridges resulting in vascular smooth muscle contraction [23]. The potent tyrosine kinase inhibitor, genistein [24] significantly

inhibited contractile response to sodium fluoride, a direct G-protein activator, in rat aorta [16]. SOV, a selective inhibitor of tyrosine phosphatase, by maintaining phosphorylation of tyrosine residues [25–30], may increase the activity of G-proteins thereby favouring vasoconstriction. By contrast, genistein and SOV had no significant effect on MLCK [17], protein kinase-C [16, 24] and protein kinase-A [24] activities. The inhibitory effect of genistein on agonist-induced contraction is still present in calcium-free medium [16]. Furthermore, the calcium channel blocker verapamil has no effect on the potentiation of agonist-induced contraction by phenylarsine oxide, an inhibitor of tyrosine phosphatase [31]. This suggests that calcium released from intracellular stores rather than calcium influx might, at least in part, mediate the vasoconstrictor effect of tyrosine kinase.

Acetylcholine causes vasodilatation through activation of eNOS and NO release. The present study provides circumstantial evidence suggesting that inhibition of eNOS through tyrosine phosphorylation may account for the inhibitory effect of SOV on acetylcholine-induced relaxation.

Previous reports have shown that both tyrosine-phosphatase inhibitors, SOV and phenylarsine oxide, caused endothelium-dependent vasodilatation in pig coronary and in rabbit carotid arteries respectively [4, 15]. In this study, only a moderate endothelium-dependent vasodilatation of rat pulmonary arteries in response to increasing concentrations of SOV was found. Pretreatment with L-NAME, however, greatly increased the vasodilator response to SOV, that was fully attenuated by glibenclamide. This suggests that NO exerts an inhibitory effect on a glibenclamide-sensitive vasodilator that can be activated by SOV. This vasorelaxing factor most likely derives from the endothelium, as PA rings, without endothelium failed to relax in response to SOV after inhibition of NO synthesis by L-NAME. As glibenclamide only inhibited the L-NAME-induced vasorelaxing response to SOV in PA rings with endothelium, the authors suggest that the putative endothelial-derived relaxing factor implicated in this study is one of the EDHF(s). To date the release of EDHF, induced by acetylcholine, has been seen only after inhibition of eNOS and cyclooxygenase [3, 11]. In coronary arteries, EDHF is likely to act through the opening of  $K_{Ca}$  channels rather than  $K_{ATP}$  channels [11, 12]. In the present study, the vasodilator effect of SOV, which is seen only in the presence of L-NAME, suggests that inhibition of NO synthesis favours the release of a vasodilator substance sensitive to glibenclamide. This compensatory mechanism may play an important role in vascular disorders characterized by impairment of NO synthesis and/or release.

In conclusion, it has been demonstrated that inhibition of tyrosine phosphatase increases the vaso-contractile response to phenylephrine and reduces endothelium-dependent relaxation to acetylcholine. Surprisingly, sodium orthovanadate also causes vasodilatation, which is probably due to the release of endothelium-derived hyperpolarizing factor, an effect

which can only be observed during inhibition of nitric oxide synthesis.

## References

1. Dinh-Xuan AT. Endothelial modulation of pulmonary vascular tone. *Eur Respir J* 1992; 5: 757–762.
2. Lévy M, Sabry S, Mercier JC, Dinh-Xuan AT. Rôles des facteurs vasoactifs synthétisés par l'endothélium dans l'hypertension artérielle pulmonaire. *Arch Pédiatr* 1997; 4: 271–277.
3. Galand CJ, Plane F, Kemp BK, Cocks TM. Endothelium-dependent hyperpolarization: a role in the control of vascular tone. *Trends Pharmacol Sci* 1995; 16: 23–30.
4. Fleming I, Bauersachs J, Fisslthaler B, Busse R.  $Ca^{2+}$ -independent activation of the endothelial nitric oxide synthase in response to tyrosine phosphatase inhibitors and fluid shear stress. *Circ Res* 1998; 82: 686–695.
5. Corson MA, James NL, Latta SE, Nerem RM, Berk BC, Harison DG. Phosphorylation of endothelial nitric oxide synthase in response to fluid shear stress. *Circ Res* 1996; 79: 984–991.
6. Ayajiki K, Kindermann M, Hecker M, Fleming I, Busse R. Intracellular pH and tyrosine phosphorylation but not calcium determines shear stress-induced nitric oxide production in native endothelial cells. *Circ Res* 1996; 78: 750–758.
7. García-Cardena G, Fan G, Stern DF, Lin J, Sessa WC. Endothelial nitric oxide synthase is regulated by tyrosine phosphorylation and interacts with caveolin-1. *J Biol Chem* 1996; 271: 27237–27240.
8. Standen NB, Quayle JM, Davies NW, Brayden JE, Huang Y, Nelson MT. Hyperpolarizing vasodilators activate ATP-sensitive  $K^+$  channels in arterial smooth muscle. *Science* 1989; 245: 177–180.
9. Brayden JE. Membrane hyperpolarization is a mechanism of endothelium-dependent cerebral vasodilation. *Am J Physiol* 1990; 259: H668–H673.
10. Brayden JE. Hyperpolarization and relaxation of resistance arteries in response to adenosine diphosphate. Distribution and mechanism of action. *Circ Res* 1991; 69: 1415–1420.
11. Nishikawa IY, Step DW, Chilian WM. In vivo location and mechanism of EDHF-mediated vasodilatation in canine coronary microcirculation. *Am J Physiol Heart Circ Physiol* 1999; 277: H1252–H1259.
12. Widmann MD, Weintraub NL, Fudge JL, Brooks LA, Dellsperger KC. Cytochrome P-450 pathway in acetylcholine-induced canine coronary microvascular vasodilation *in vivo*. *Am J Physiol Heart Circ Physiol* 1998; 274: H283–H289.
13. Li PL, Zou AP, Campbell WB. Regulation of potassium channels in coronary arterial smooth muscle by endothelium-derived vasodilators. *Hypertension* 1997; 29: 262–267.
14. Lévy M, Souil E, Sabry S, *et al.* Maturation changes of endothelial vasoactive factors and pulmonary vascular tone at birth. *Eur Respir J* 2000; 15: 158–165.
15. Nakaike R, Shimokawa H, Owada MK, *et al.* Vanadate causes synthesis of endothelium-derived NO *via* pertussis toxin-sensitive G protein in pigs. *Am J Physiol Heart Circ Physiol* 1996; 271: H296–H320.
16. Abebe W, Agrawal DK. Role of tyrosine kinase in

- norepinephrine-induced contraction of vascular smooth muscle. *J Cardiovasc Pharmacol* 1995; 26: 153-159.
17. Jin NJ, Siddiqui RA, English D, Rhoades RA. Communication between tyrosine kinase pathway and myosin light chain kinase pathway in smooth muscle. *Am J Physiol Heart Circ Physiol* 1996; 271: H1348-H1355.
  18. Di Salvo J, Steusloff A, Semenchuck L, Satoh S, Kolquist K, Pfitzer G. Tyrosine kinase inhibitors suppress agonist-induced contraction in smooth muscle. *Biochem Biophys Res Commun* 1993; 190: 968-974.
  19. Tsuda T, Kawahara Y, Shii K, Koide M, Ishida Y, Yokoyama M. Vasoconstrictor-induced protein-tyrosine phosphorylation in cultured vascular smooth muscle cells. *FEBS Lett* 1991; 285: 44-48.
  20. Koide M, Kawahara Y, Tsuda T, Ishida Y, Shii K, Yokoyama M. Endothelin-1 stimulates tyrosine phosphorylation and the activities of two mitogen-activated protein kinases in cultured vascular smooth muscle cells. *J Hypertens* 1992; 10: 1173-1182.
  21. Molloy CJ, Taylor DS, Weber H. Angiotensin-II stimulation of rapid protein tyrosine phosphorylation and protein kinase activation in rat aortic smooth muscle cells. *J Biol Chem* 1993; 268: 7338-7345.
  22. Hollenberg MD. Tyrosine kinase pathways and the regulation of smooth muscle contractility. *Trends Pharmacol Sci* 1994; 15: 108-114.
  23. Kamm KE, Stull JT. The function of myosin and myosin light chain kinase phosphorylation in smooth muscle. *Annu Rev Pharmacol Toxicol* 1985; 25: 593-620.
  24. Akiyama T, Ishida J, Nakgawa S, *et al.* Genistein, a specific inhibitor of tyrosine-specific protein kinases. *J Biol Chem* 1987; 262: 5592-5595.
  25. Laniyonu A, Saiffeddine M, Ahmad S, Hollenberg MD. Regulation of vascular and gastric smooth muscle contractility by pervanadate. *Br J Pharmacol* 1994; 113: 403-410.
  26. Heffetz D, Bushkin I, Dror R, Zick Y. The insulinomimetic agents H<sub>2</sub>O<sub>2</sub> and vanadate stimulate tyrosine phosphorylation in intact cells. *J Biol Chem* 1990; 265: 2896-2902.
  27. Faure R, Bquiran G, Bergeron JJ, Posner BI. The dephosphorylation of insulin and epidermal growth factor receptors. Role of endosome associated phosphotyrosine phosphatase(s). *J Biol Chem* 1992; 267: 11215-11221.
  28. Alexander DR. The role of phosphatases in signal transduction. *News Biol* 1990; 2: 1049-1062.
  29. Atkinson TP, Lee CW, Rhee SG, Hohman RJ. Orthovanadate induces translocation of phospholipase C- $\gamma$ 1 and - $\gamma$ 2 in permeabilized mast cells. *J Immunol* 1993; 151: 1448-1455.
  30. Chao W, Liu H, Hanahan DJ, Olson MS. Protein tyrosine phosphorylation and regulation of the receptor for platelet-activating factor in rat Kupffer cells. Effect of sodium vanadate. *Biochem J* 1992; 288: 777-784.
  31. Savineau JP, Gonzalez De La Fuente P, Marthan R. Effect of modulators of tyrosine kinase activity on agonist-induced contraction in the rat pulmonary vascular smooth muscle. *Pulmonary Pharmacol* 1996; 9: 189-195.