Supplemental material

Preclinical validation and imaging of Wnt/beta-catenin-induced lung repair in human 3D lung tissue cultures

Franziska E. Uhl¹, Sarah Vierkotten¹, Darcy E. Wagner¹, Gerald Burgstaller¹, Rita Costa¹, Ina Koch², Michael Lindner², Silke Meiners¹, Oliver Eickelberg¹, and Melanie Königshoff¹

¹ Comprehensive Pneumology Center, Helmholtz Center Munich, Ludwig Maximilians University Munich, University Hospital Grosshadern, Member of the German Center for Lung Research (DZL), Munich, Germany

²Asklepios Clinics, Gauting, Germany



Supplemental Figure 1: Schematic overview of the generation of 3D *ex vivo* tissue cultures (3D-LTC) from murine and human samples. Lung tissue was filled with agarose and sliced with a vibratome to 300 (murine) or 500 µm sections (human). A detailed protocol is provided in the Material and Methods section.



Supplemental Figure 2: Characterization of cell viability in 3D-LTC over culture time. a) WST-1 conversion of the cells in 3D-LTC from control and emphysematous C57BI/6 mice cultivated for five days. n=6-14. *: p<0.05. b) Quantification of the cleaved caspase 3 positive stained cells in 3D-LTC from healthy C57BI/6 mice at different time points of cultivation. *: significant to day1, #: significant to other time point, *: p<0.05, **: p<0.01, ***: p<0.001. c) H&E staining of 3D-LTC from healthy C57BI/6 mice during cultivation. Black arrows show fragmented nucleii. Scale bars 200 µm and 50 µm.



Supplemental Figure 3: Lung structure in murine 3D-LTC over culture time. Immunofluorescence staining of collagen I in 3D-LTC from healthy and emphysematous C57BI/6 mice at day 5 of cultivation. Scale bars 50 µm.



Supplemental Figure 4: Time course of elastase-induced emphysema. H&E staining of 3D-LTC at day 0 from healthy and emphysematous C57Bl/6 mice at different time points after elastase or PBS instillation. Scale bar 500 µm.



Supplemental Figure 5: Mitochondrial activity upon LiCI and CT treatment in 3D-LTC and 2D cell culture. WST-1 conversion of the cells in 3D-LTC from healthy (a) and emphysematous (b, c) C57BI/6 mice treated with different concentrations of LiCI and CT for different periods. d) WST-1 conversion of the MLE12 epithelial cells and MHS macrophages treated with different concentrations of LiCI for 24 h compared to control (=100%). n=3-4. *: p<0.05, **: p<0.01, ***: p<0.001.



Supplemental Figure 6: Assessment of macrophage speed and length in healthy and emphysematous 3D-LTC by 4D confocal live tissue imaging. Quantification of track length (a) and track speed (b) of macrophages in 3D-LTC from healthy and emphysematous MacGreen animals with LiCI treatment for 48 h. Box plots of track length (c) and track speed (d). ***: p<0.001. Quantification derived from Video 3.



Supplemental Figure 7: *Mmp12* and *Eln* level upon Wnt/beta-catenin activation by 2 μM CT in murine 3D-LTC. Gene expression analysis by qPCR in 3D-LTC from emphysematous C57BI/6 mice treated with CT for different periods. n=3-4. *: p<0.05, **: p<0.01.



Supplemental Figure 8: Characterization of cell viability in human 3D-LTC over culture time. a) WST-1 conversion of the cells in 3D-LTC from non-COPD and COPD patients cultivated for five days. n=4-8. *: p<0.05. b) Representative Live/Dead confocal images of the different planes of 3D-LTC from non-COPD and COPD patients at the indicated time points. Ortho views of the top, middle, and bottom plane. Healthy cells are indicated by green cytoplasm (arrows) and dead cells by red nucleus (arrowheads).



Supplemental Figure 9: Lung structure in patient-derived 3D-LTC over culture time. IF staining of collagen I of 3D-LTC from non-COPD and COPD patients at day 5 of cultivation. Scale bars 50 μm.

	COPD	Non-COPD
	(n = 11)	(n = 14)
Patient age at time of surgery (years)	66 ± 7.8	62.1 ± 8.6
Gender (M:F)	7:4 (63.6%/36.4%)	7:7 (50%/50%)
Smoking status		
Current smoker	8 (72.7%)	5 (35.7%)
Former smoker	2 (18.2%)	5 (35.7%)
Never	0 (0%)	2 (14.2%)
Unknown	1 (9.1%)	2 (14.2%)
Physiology		
%FEV ₁	74.2 ± 13.6	107.1 ± 15.1
FEV ₁ /FVC %	64.7 ± 5.6	80.1 ± 5.6
%DLCO/VA	63.7 ± 25.3 [†]	74.2 ± 17.6 [†]
COPD stage (GOLD Guidelines)		
I	5 (45.5%)	
II	6 (54.5%)	
Cancer type		
Adenocarcinoma	5 (45.5%)	10 (71.4%)
Squamous cell carcinoma	2 (18.2%)	3 (21.4%)
Other adenocarcinomas*	4 (36.4%)	1 (7.1%)

Supplemental Table 1. Clinical characteristics of patients included in this study

Data are given as mean ± SD or as number of patients (%) * metastasis from organ other than the lung † three unknown values

Gene		Sequence 5' - 3'
ACTA2	fw	CGAGATCTCACTGACTACCTCATGA
	rv	AGAGCTACATAACACAGTTTCTCCTTGA
AXIN2	fw	AGAAATGCATCGCAGTGTGAAG
	rv	GGTGGGTTCTCGGGAAATG
COL 1 A 1	fw	CAAGAGGAAGGCCAAGTCGAG
COLIAI	rv	TTGTCGCAGACGCAGATCC
DKK2	fw	GATGTCACATATAAAAGGGCATGAAG
	rv	AATGACGAGCACAGCAAAACC
HOPX	fw	GCCCCACAGAGGACCAGGTG
	rv	GCTTGGTTAAGCGGAGGAGAG
HPRT	fw	AAGGACCCCACGAAGTGTTG
	rv	GGCTTTGTATTTTGCTTTTCCA
MMP12	fw	TGCTGATGACATACGTGGCA
	rv	AGGATTTGGCAAGCGTTGG
NKD1	fw	CACCCTGTATGACTTTGACAACAAC
	rv	CAGAGGAGTCCACCACCTCATAG
PDPN	fw	GAGAAAGATGGTTTGTCAACAGTG
	rv	GGCGTAACCCTTCAGCTCT
SETDC	fw	GCCCAGTGCACCTGAAACGC
SFIFU	rv	TCTCCAGAACCATCTCCGTGTGT

Supplemental Table 2. Human primers used for qPCR

Gene		Sequence 5' - 3'
f ^r Axin2 r	fw	AGCAGAGGGACAGGAACCA
	rv	CTGAACCGATTCATGACCAC
Col1a1	fw	AGCTTTGTGGATACGCGGACT
	rv	TCGTACTGATCCCGATTGCA
	fw	GAGATCGCAACCATGGTCACT
DKKZ	rv	GGGTCTCCTTCATGTCCTTTTATATG
Elp	fw	GGCGTCTTGCTGATCCTCT
	rv	ATAATAGACTCCACCGGGAACT
Eafr	fw	ACACAGATAGCTCCGAAGACGTTGT
rgiiz	rv	CCCAGCCGGACAGCGGAACT
—	fw	GGTGTAGCACAACTTCCAATTACG
Fn	rv	GGAATTTCCGCCTCGAGTCT
Норх	fw	TCTCCATCCTTAGTCAGACGC
	rv	GGGTGCTTGTTGACCTTGTT
Hprt	fw	CCTAAGATGAGCGCAAGTTGAA
	rv	CCACAGGACTAGAACACCTGCTAA
Mmn12	fw	TGTACCCCACCTACAGATACCTTA
winp 12	rv	CCATAGAGGGACTGAATGTTACGT
Nkd1	fw	TGTTCTCATCCACGCAATGG
	rv	GAGCCCCACTCAGGTTCCA
Ddan	fw	ACAGGTGCTACTGGAGGGCTT
Papn	rv	TCCTCTAAGGGAGGCTTCGTC
Sftpc	fw	AGCAAAGAGGTCCTGATGGA
	rv	GAGCAGAGCCCCTACAATCA
11/22	fw	AGCCCTGATGAACCTTCACAAC
VV I ITZ	rv	TGACACTTGCATTCTTGTTTCAAG

Supplemental Table 3. Murine primers used for qPCR

Nama	Source	Manufacturar	Ordor pr	Dilution	Size
Name		Manufacturer	Order III.		[kDa]
β-actin	Mouse	Sigma-Aldrich; Taufkirchen	A3854	1:50,000	42
		Germany			12
β-catenin, active	Mouse	Millipore; Billerica, MA, USA	05-665	1:500	92
Elastin	Mouse	Millipore; Billerica, MA, USA	MAB2503	1:1,000	68
Норх	Rabbit	Santa Cruz; Santa Cruz, CA,	sc-30216	1:1,000	13
		USA			
Podoplanin	Goat	R&D Minneapolis, MN, USA	AF3244	1:1,000	40
Podoplanin	Rat	Acris; Herford, Germany	AM01133PU-N	1:1,000	37

Supplemental Table 4. Antibodies for Western Blot

Supplemental Table 5. Antibodies for immunofluorescence

Name	Source	Manufacturer	Order nr.	Dilution
Aquaporin 5 Ra	Rabbit	Calbiochem, Millipore; Billerica, MA,	178615	1:100
	Rabbit	USA	170010	
CD45	Rat	BD; San Jose, CA, USA	553076	1:500
Cleaved	Dabbit	Cell Signalling Technology; Boston,	0661	1.250
caspase3	Rappil	MA, USA	9001	1.230
Collagen 1	Rabbit	Rockland; Gilbertsville, PA, USA	600-401-103	1:100
E-cadherin	Mouse	BD; San Jose, CA, USA	610181	1:100
Podoplanin	Goat	R&D Minneapolis, MN, USA	AF3244	1:100
Podoplanin Ra	Det	Acris; Herford, Germany	AM01133PU-	1:100
	Ral		Ν	
Sftpc, pro-	Rabbit	Abcam; Cambridge, UK	ab40879	1:500
Sftpc, pro-	Rabbit	Millipore; Billerica, MA, USA	AB3786	1:500

Supplemental Videos

Video 1: Ciliated cell of a murine 3D-LTC at day seven. Cilia are still beating, demonstrating cell viability and functionality.

Video 2: 4D confocal live tissue imaging of a 3D-LTC from an emphysematous TCF/LEF-GFP animal showing increases in GFP positive cells following 10 mM LiCI treatment.

Video 3: 4D confocal live tissue imaging of 3D-LTC from healthy MacGreen animals treated with ctrl. (left panel) or 10 mM LiCI (right panel) for 48 h showing subjective decreases in macrophage speed and distance travelled following LiCI treatment (quantitative characterization in Supplemental Figure 6).

Video 4: 4D confocal live tissue imaging of 3D-LTC from emphysematous MacGreen animals treated with ctrl. (left panel) or 10 mM LiCl (right panel) for 48 h showing subjective decreases in macrophage speed and distance travelled following LiCl treatment (quantitative characterization in Supplemental Figure 6).